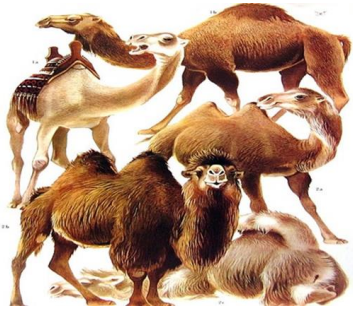


Differential splicing: an effective way to expand molecular diversity of caseins and their ability to generate bioactive peptides



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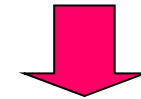
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Milk: a complete and complex food

Constant
physical
aspect

Hide a large variability in
composition between
species

a paradox

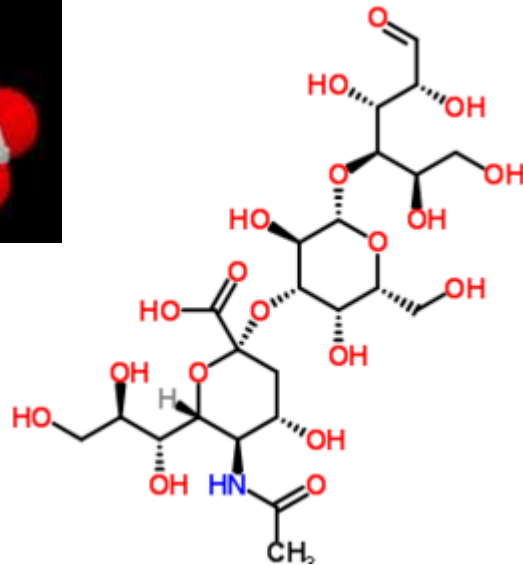
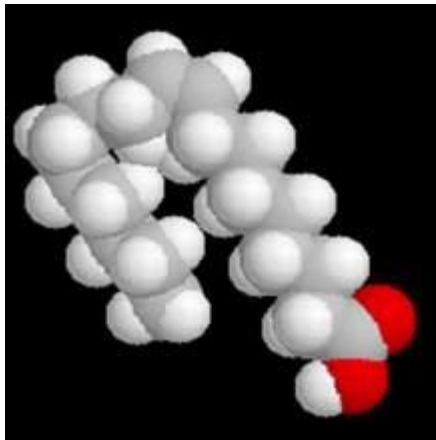


“Designed” to fulfil **specific** requirements
and provide optimal growth and
development to mammalian **offsprings**



Milk is much more than a complete food

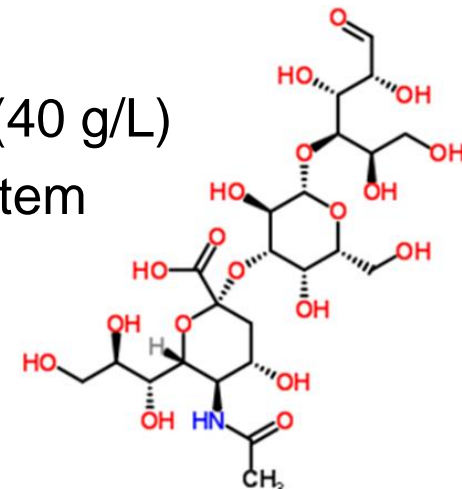
Milk is a source of bioactive components, including long chain fatty acids, complex oligosaccharides and proteins



HMOs: the third most abundant solid component of human milk



- 10 g/L in mature milk after lactose (70 g/L) and lipids (40 g/L)
- Confer protection on the newborn whose immune system is functionally immature
- Prebiotics, promoting growth of beneficial bacteria,
- Protect against pathogenic infection
- Stimulate maturation of the immune system

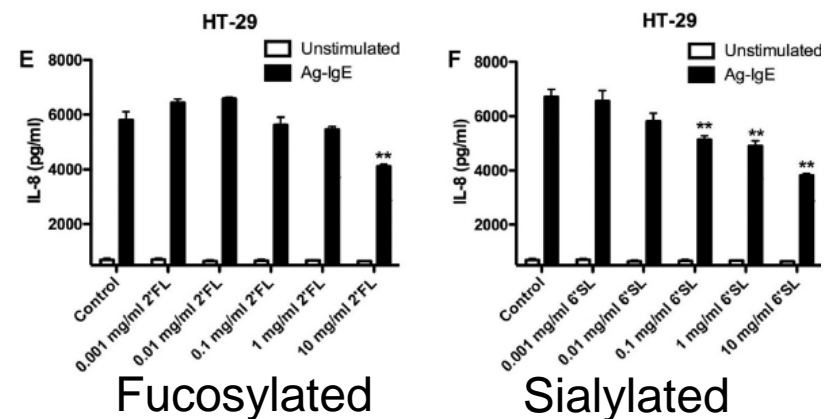


Fucosylated 40%
Sialylated 13%

Human Milk Oligosaccharides Attenuate Antigen–Antibody Complex Induced Chemokine Release from Human Intestinal Epithelial Cell Lines

Sehrish Zehra, Ibrahim Khambati, Megan Vierhout, M. Firoz Mian, Rachael Buck, and Paul Forsythe

Vol. 83, Nr. 2, 2018 • Journal of Food Science



Oligosacharrides in camel milk



J. Dairy Sci. 93:5572–5587
doi:10.3168/jds.2010-3151
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Chemical characterization of the oligosaccharides in Bactrian camel (*Camelus bactrianus*) milk and colostrum

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J. Dairy Sci. 103:72–86
<https://doi.org/10.3168/jds.2019-16710>
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Electrospray ionization mass spectrometry characterization of ubiquitous minor lipids and oligosaccharides in milk of the camel (*Camelus dromedarius*) and their inhibition of oxidative stress in human plasma

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Milk proteins: a source of biologically active peptides



β -casomorphins (BCMs), derived from β -casein, first identified in bovine milk

(Brantl & Teschemacher, 1979)

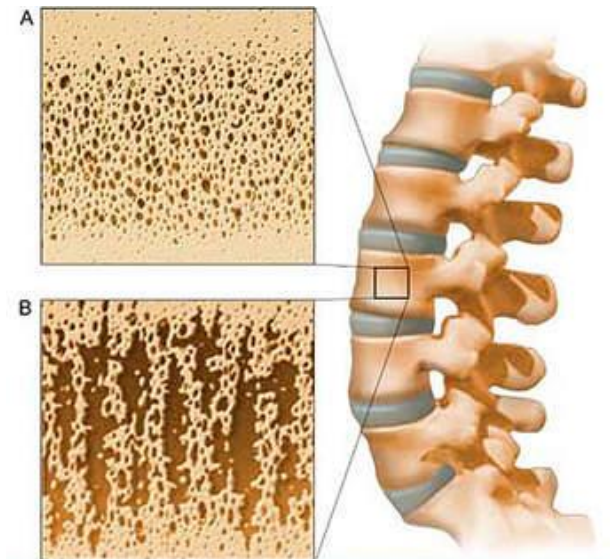
recently shown to occur naturally in breast milk (Enjapoori *et al.*, 2019)

Signal peptide

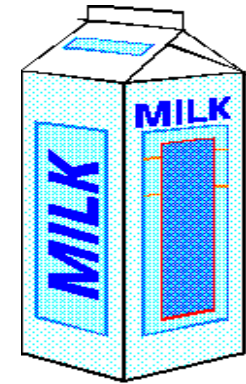
10	20	30	40	50	
MKVLILACLVALALA	RELEE	LNVPGEIVES	LSSSEESITR	INKKIEKFQS	
60	70	80	90	100	
EEQQQTEDEL	QDKIHPFAQT	QSLV YFPF GGP	IHNSLPQNIP	PLTQTPVVVP	BCM5-11
110	120	130	140	150	
PFLQPEVMGV	SKVKEAMAPK	HKEMPF'PKYP	VEPFTESQSL	TLTDVENLHL	
160	170	180	190	200	
PLPLLQSWMH	QPHQPLPPTV	MFPPQSVLSL	SQSKVLPVPQ	KAVPYPQRDM	
210	220				
PIQAFLLYQE	PVLGPVRGPF	PIIV			

Therapeutic effects of milk-derived peptides in several health disorders

Hypertension
cardiovascular disease
obesity
osteoporosis
dental caries
gastrointestinal diseases
ageing, ... and others

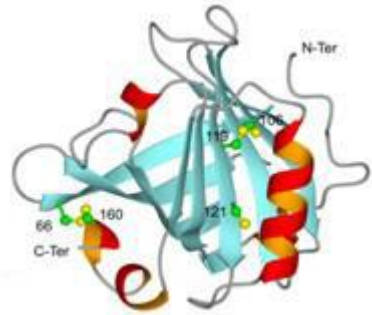


Milk proteins



Soluble fraction
(Whey proteins)

Colloidal fraction



20%

β -lactoglobulin

α -lactalbumin

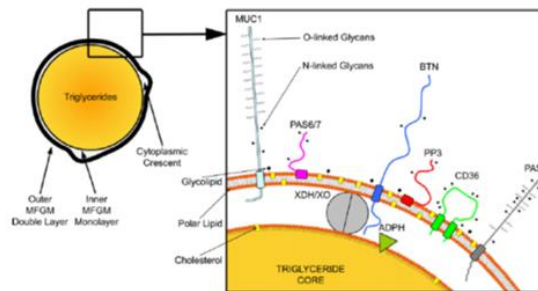
Lactoferrin, ...

(Globular tertiary structure)

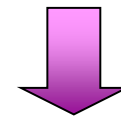
Milk Fat Globule Membrane

> 80 proteins (<1%)

(Butyrophilin, adipophilin, xanthine oxidase, FABP, lactadherin, ...)



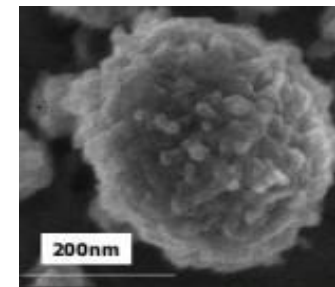
(Dewettinck *et al.*, 2008)



80%

Caseins

(random coiled, organised in Micelles)



(Dagleish *et al.*, 2004)

The bulk of the protein fraction in milk corresponds to the products of only 6 genes

Coding for: 4 caseins (α s1, α s2, β and κ)
2 main whey proteins (β -lactoglobulin & α -lactalbumin)

95% of the protein fraction in cattle




Several tens or even hundreds of proteins in the remaining 5%

A fairly simple system, regarding the main milk proteins, but this milk proteome is much more complex with:

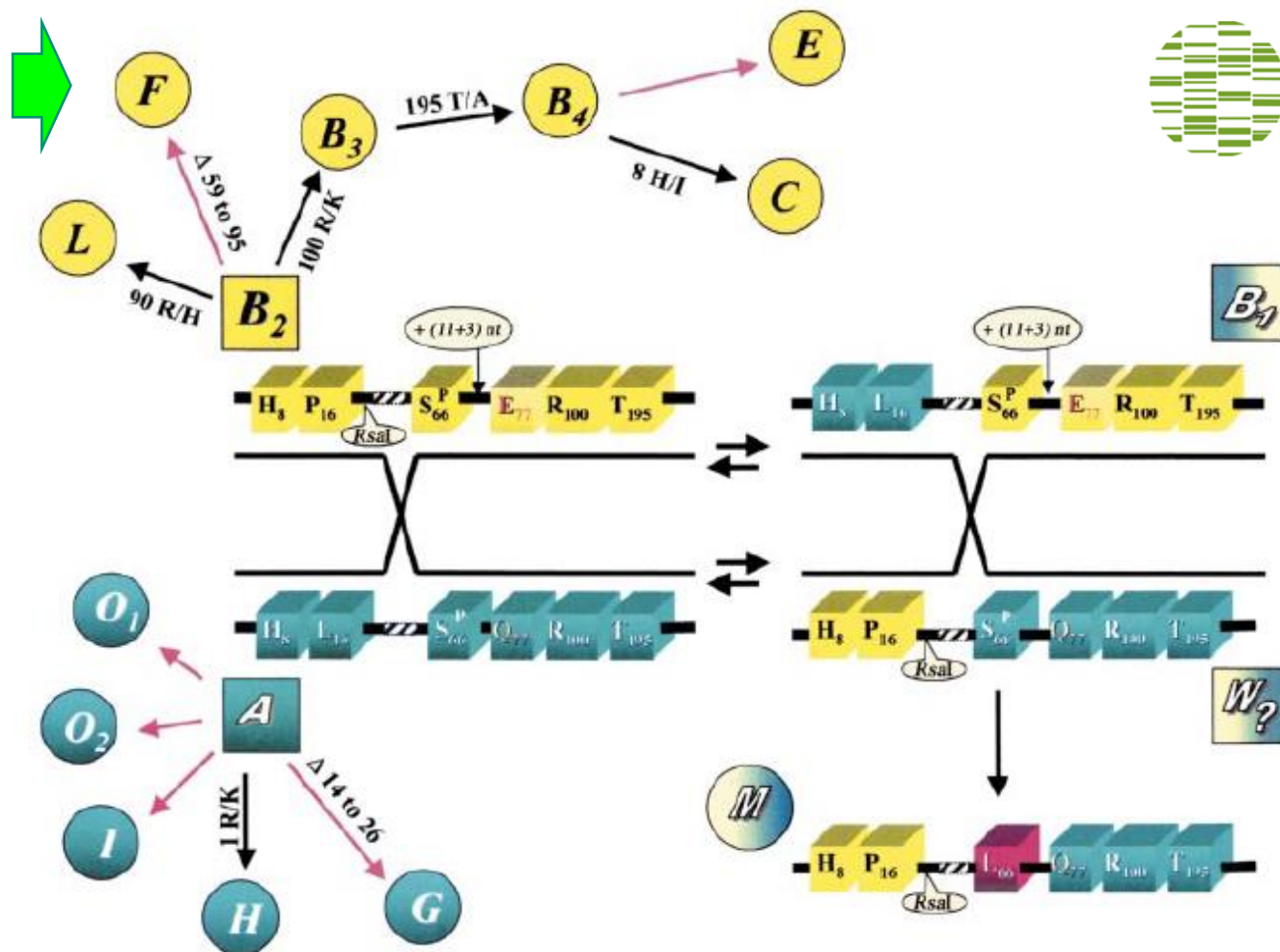
- **genetic variants**
- low-abundance gene products arising from **differential splicing**
- isoforms due to **post-translational modifications** such as **phosphorylation**, glycosylation and proteolytic processing.

up to 100 molecules, derived from the 6 major milk proteins

Genetic variants of the main milk proteins

	CSN1S1 (α s1-casein)	CSN1S2 (α s2-casein)	CSN2 (β -casein)	CSN3 (κ -casein)	β-lactoglobulin	α-lactalbumin
	9 variants A to I	4 variants A to D	12 variants A ¹ , A ² , A ³ , B to J	15 variants A, B, B2, C to E (F1, F2, G1, G2 and H to L)	13 variants A, B, B*, C, D and Dr, E to J and W	4 variants A to D
	15 variants (+ 3 null alleles)	7 variants (+1 null allele)	6 variants (+ 3 null alleles)	17 variants	2 putative variants (uncharacterized so far)	2 Variants
	10 variants	8 variants	5 variants	3 variants	3 variants	2 variants

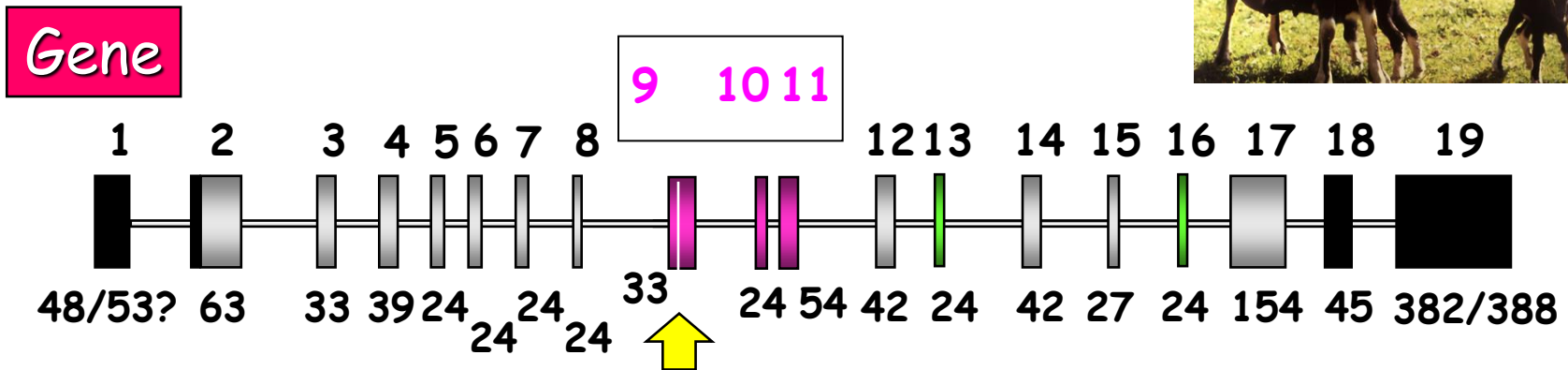
Genetic polymorphisms at the *CSN1S1* locus in the goat species



Allele F: A "frame shift" mutation

a multiplicity of transcripts (proteins)

a reduced expression



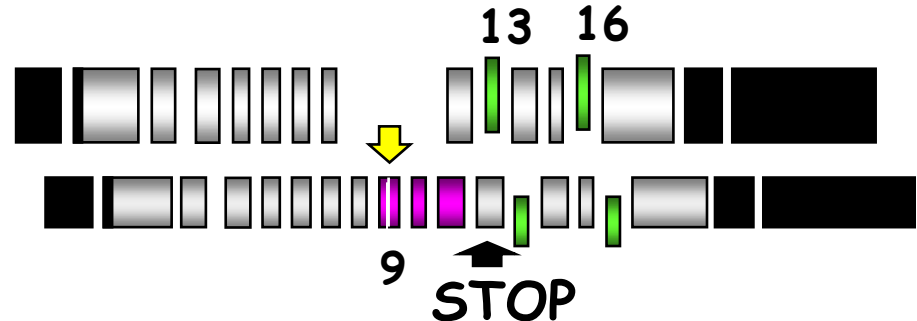
Leroux *et al.*, 1992

α_{s1} -CasA



mRNA

α_{s1} -CasF



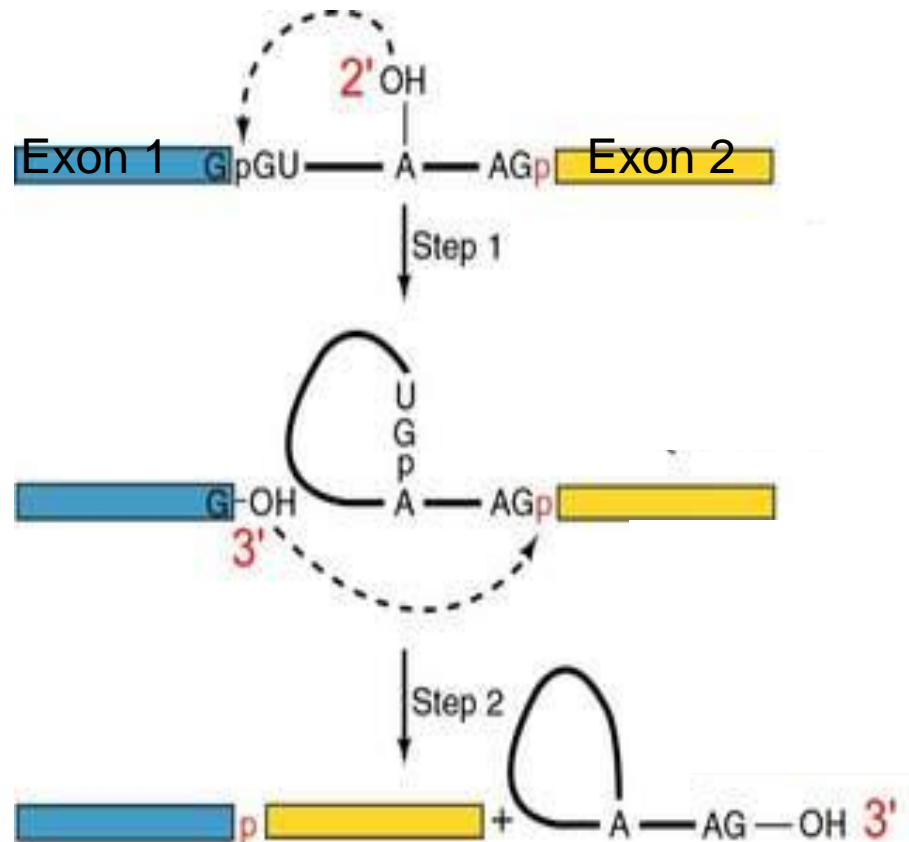
α_{s1} -CasF → 9 different transcripts as many different peptide chains

Occurrence of splicing variants due to cryptic splice site usage and exon skipping

Exon skipping (ES): loss of exon during the course of pre-mRNA maturation (assembly of exonic sequences and removal of introns)

Cryptic splice site usage (CSS): loss or gain of nucleotides (intron or exon sequences)

Quality control mechanisms

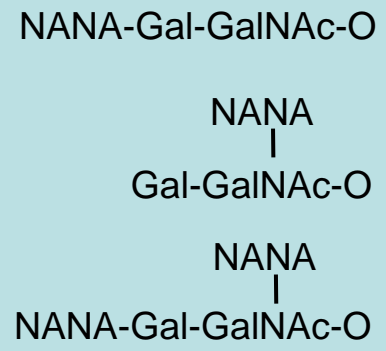


Events first reported in small ruminants: CSN1S in the goat species

Post-translational modifications contribute to increase molecular diversity

Glycosylation: κ -casein (O-glycosylation on up to 6 Thr residues)

MMKSFFLVVTILALTLPLGA QEQNQEQP I RCEKDERFFSDKIAKYIPIQY
 VLSRYPSYGLNYYQQKPVALINNQFLPYPYAKPAAVRSPAQILQWQVLSNT
 VPAKSCQAQPTTMARHPHPHLS **F-MA** I PPKKNQDKTEIP **T**INTIA **SGEPTST**
 PT**T**EAVES**T**VA**TLE****D****SPE** VIESPPEINTVQV**T**STAV



Phosphorylation: α s2-casein (theoretically up to 16P on Ser/Thr residues)

MKFFIFTCLLAVALA KN**TME**HV **SSSEE** SII **SQE** TYKQEKNMAINP **SKE**NL
 CSTFCKEVVRNANEEEEYSIG **SSSEESAEVA** **TEE** VKITVDDKHYQKALNEIN
 QFYQKFPQYLQYLYQGPIVLNPWDQVKRNAVPITPTLNREQL **STSEE**NSKK
 TVDME **STE**VFTKKTKL **TEE**EKNRLNFLKKISQRYQKFALPQYLKTVYQHOK
 AMKPWIQPKTKVIPYVRYL

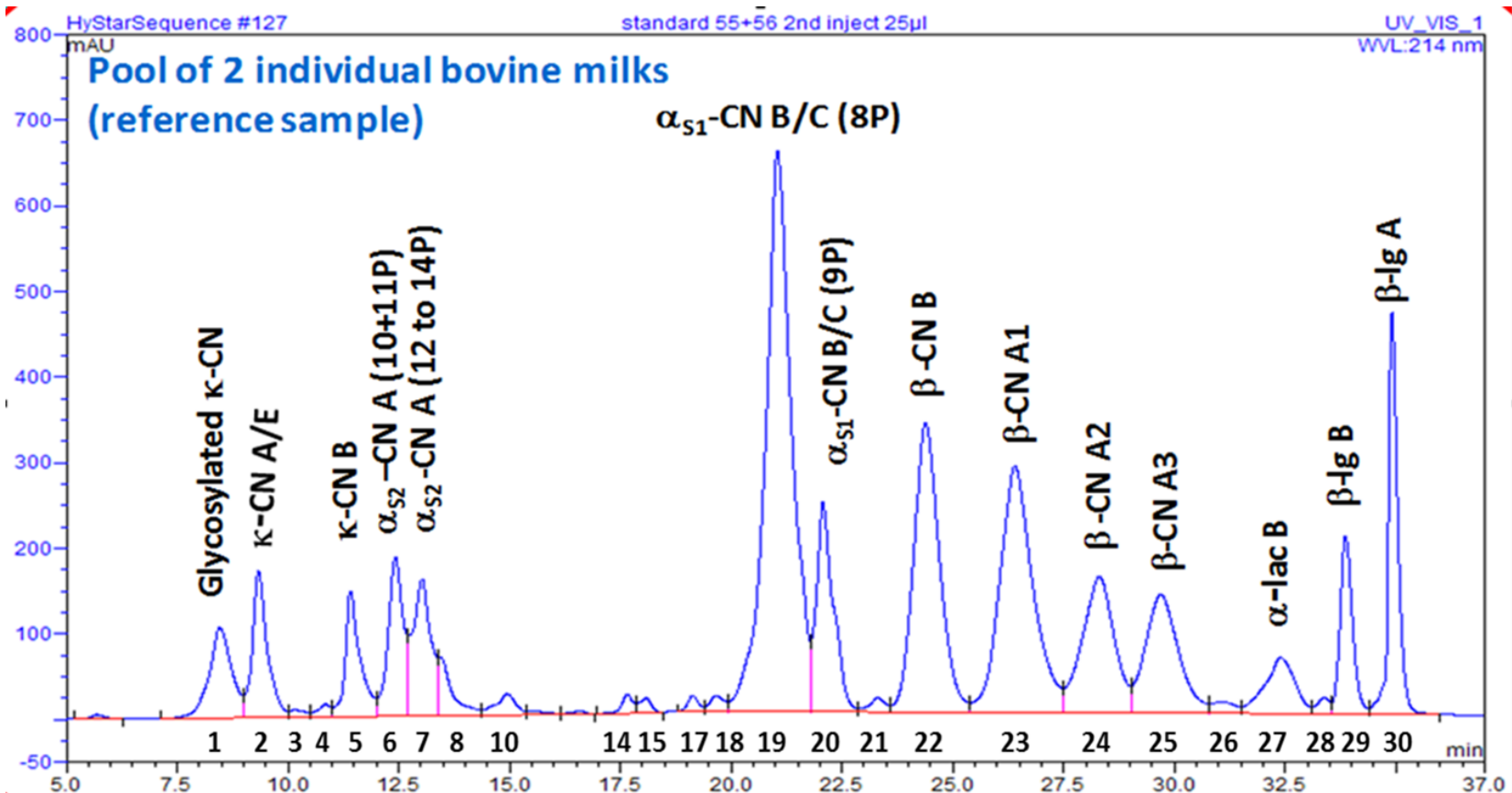
LC-MS profiling of bovine milk proteins



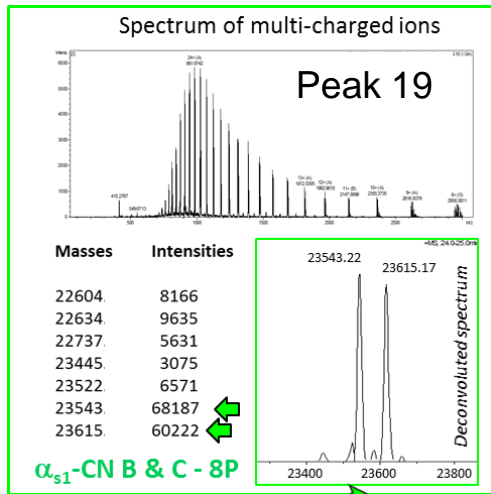
An improved LC-MS method to profile molecular diversity and quantify the six main bovine milk proteins, including genetic and splicing variants as well as post-translationally modified isoforms

Guy Miranda^{a,*}, Leonardo Bianchi^a, Zuzana Krupova^{a,1}, Philippe Trossat^b, Patrice Martin^{a,*}

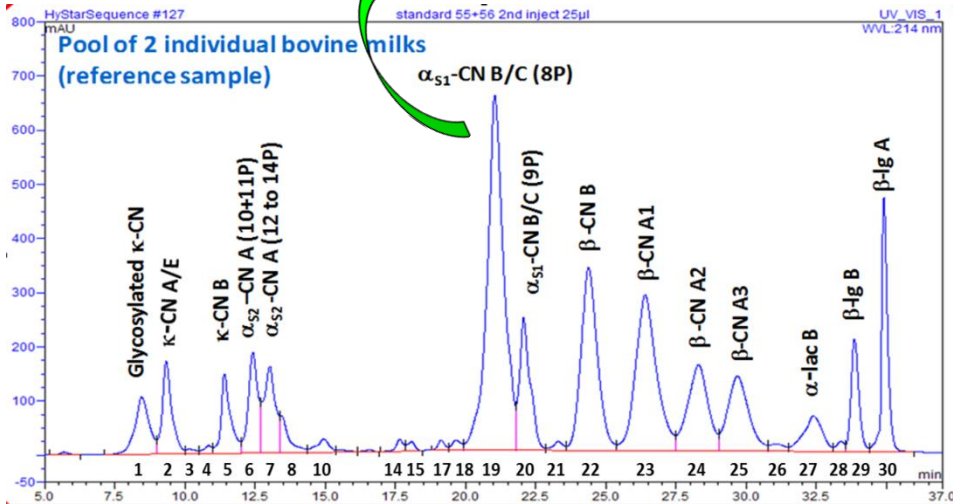
^a UMR GABI, INRAE, AgroParisTech, Université Paris-Saclay, 78350 Jouy-en-Josas, France
^b ACTALIA, pôle expertise analytique, 39801 Poligny, France



LC-MS profiling of bovine milk proteins



Peak number	Retention Time	Main masses Observed	Identification	Theoretical masses	Area	Relative Area	Others	Protein family	Relative concentration
	min	Da		Da	mAU*min	%			%
1	8.46	20901.8615 20931.8947 21850.3951	κ -CN B 1P 2OG κ -CN A 1P 2OG κ -CN E 1P 3OG	20901.098 20933.054 21850.889	60.187	2.97		Glycosylated κ -CN	2,97
2	9.34	19007.3305 19037.2153	κ -CN E 1P κ -CN A 1P	19007.345 19037.371	66.752	3.30		κ -CN	3.30
3	10.15		n.a.		3.133	0.15	0.15		
4	10.84		n.a.		4.843	0.24	0.24		
5	11.41	19005.3940	κ -CN B 1P	19005.416	56.887	2.81		κ -CN	2.81
6	12.42	25148.4917 25228.5579	α s2-CN A 10 & 11P	25148.348 25228.328	74.018	3.66		α s2-CN	8,43
7	13.02	25308.4656	α s2-CN A 12P	25308.308	77.015	3.81			
8	13.43	25388.2338 25468.6767	α s2-CN A 13 & 14P	25388.288 25468.288	19.390	0.96			
9	13.89		n.a.		5.193	0.26	0.26		
10	14.95	12096.3734 12177.1687	PP5 f(1-105) A2/A3 4P PP5 A2/A3 5P	12097.294 12177.274	14.213	0.70		β -CN	0.70
11	15.58		n.a.		3.398	0.17	0.17		
12	16.80		n.a.		3.0344	0.15	0.15		
13	17.38		n.a.		1.450	0.07	0.07		
14	17.66	21997.0835 22068.8598	α s1-CN C 8P Δ e4 α s1-CN B 8P Δ e4	21996.831 22068.910	7.395	0.37		α s1-CN	0.75
15	18.07	14031.9655 14105.6761	f(80-199) α s1-CN C 1P f(80-199) α s1-CN B 1P	14035.772 14107.836	7.746	0.38			
16	18.64		n.a.		1.598	0.08	0.08		
17	19.14	22426.9941 22601.9471	α s1-CN B 8P Δ e8 Δ Q59 & Q78 α s1-CN B 7P Δ e8	22427.422 22603.703	8.347	0.41		α s1-CN	0.41
18	19.66		n.a.		9.622	0.48	0.48		
19	21.04	23542.3689 23614.8144	α s1-CN C 8P α s1-CN B 8P	23542.648 23614.712	482.326	22.84		α s1-CN	28.48
20	22.07	23624.2794 23695.6948	α s1-CN C 9P α s1-CN B 9P	23622.628 23694.692	114.171	5.64			
21	23.29		n.a.		9.975	0.49	0.49		
22	24.39	24092.5726	β -CN B 5P	24092.319	237.283	11.72		β -CN	37.91
23	26.40	24023.4898	β -CN A1 5P	24023.209	257.687	12.73			
24	28.31	23983.5180	β -CN A2 5P	23983.185	138.520	6.84			
25	29.69	23974.1909	β -CN A3 5P	23974.174	124.989	6.17			
26	31.09	11558.4804	γ 3 f(108-209) β -CN 0P	11558.612	9.201	0.45			
27	32.40	14185.8586	α -lactalbumin B	14186.084	53.580	2.64			
28	33.37	18605.2732	lactosyl- β -lactoglobulin B	18605.320	6.831	0.34		β -LG	9,49
29	33.86	18281.1176	β -lactoglobulin B	18281.208	68.804	3.40			
30	34.91	18367.2233 18691.6191	β -lactoglobulin A lactosyl- β -lactoglobulin A	18367.298 18691.300	116.472	5.75			
Total					2024.040	100.00	2.10		97,90



LC-MS profiling of Camel milk proteins

Combining different proteomic approaches to resolve complexity of the milk protein fraction of dromedary, Bactrian camels and hybrids, from different regions of Kazakhstan

Alma Ryskaliyeva^{1*}, Céline Henry², Guy Miranda¹, Bernard Faye³,
Gaukhar Konuspayeva⁴, Patrice Martin¹

¹ INRA, UMR GABI, AgroParisTech, Université Paris-Saclay, Jouy-en-Josas, France, ² INRA, MICALIS Institute, Plateforme d'Analyse Protéomique Paris Sud-Ouest (PAPPSO), Université Paris-Saclay, Jouy-en-Josas, France, ³ CIRAD, UMR SELMET, France, ⁴ Al-Farabi Kazakh State National University, Biology department, Almaty, Kazakhstan



2018



peak I: κ -CN

peak III: WAP

peaks IV & V: α s1-CN

peak VI: α -LAC,

peak VII: α s2-CN

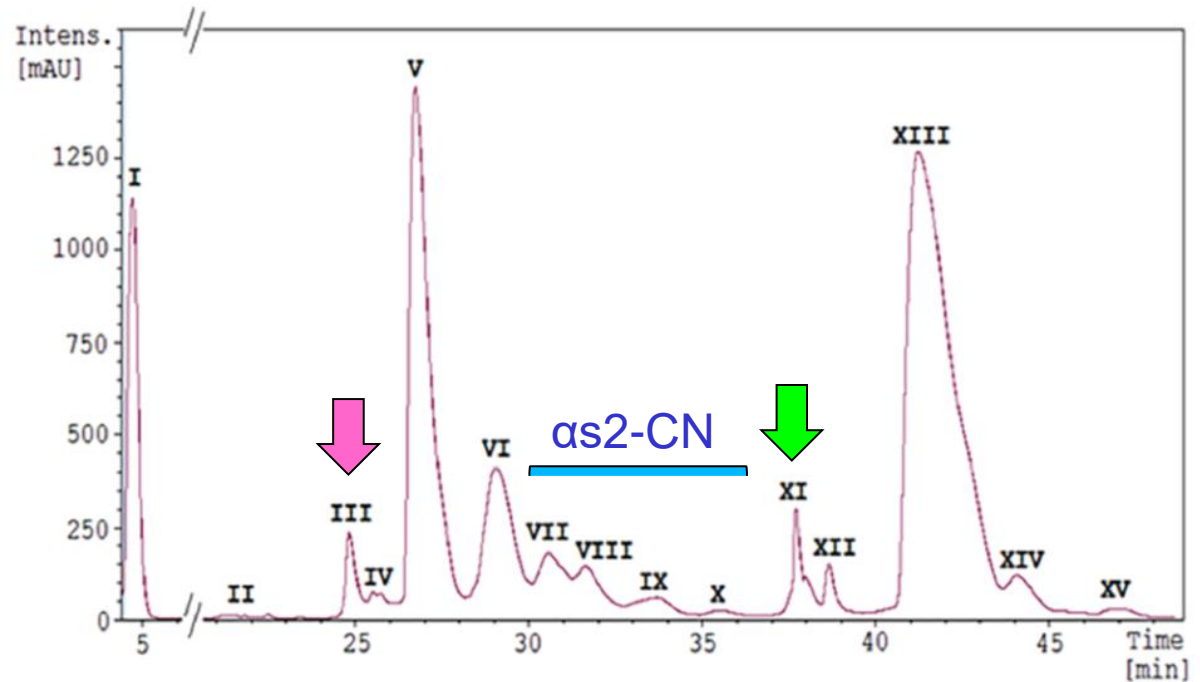
peaks VIII, IX, & X: α s2-CN

peak XI: PGRP

peak XII: CSA/LPO

peaks XIII & XIV: β -CN

peak XV: γ 2-CN.



Peak	Ret. Time, min	Observed M _r , Da	Theoretical M _r , Da	Protein description	UniProt/ NCBI GenBank Accession number	Intensity		
I	4.50	21,157	21,158	κ-CN A, 1P, (GaN-Ga-SA2)x3*, pyro-E		1,361		
		21,184	21,182	κ-CN B, 0P, (GaN-Ga-SA2)x3 + (GaN-Ga-SA2)x2**, pyro-E		5,810		
II	18.61	18,210	18,210	κ-CN B, 0P ?	L0P304	161		
		18,236	18,235	κ-CN A, 0P, pyro-E	P79139	72		
III	24.32	12,564	12,564	WAP, 0P	P09837	1,756		
		12,644	12,644	WAP, 1P		1,575		
IV	24.97	23,878	23,878	α _{s1} -CN A—short isoform (Δex 16 and Δex 13'), 4P		242		
V	26.23	24,547	24,547	α _{s1} -CN C -short isoform (Δex 16), 5P, splice variant (ΔQ83)		4,885		
		24,627	24,627	α _{s1} -CN C—short isoform (Δex 16), 6P, splice variant (ΔQ83)		21,606		
		24,707	24,707	α _{s1} -CN C—short isoform (Δex 16), 7P, splice variant (ΔQ83)		6,990		
		24,675	24,675	α _{s1} -CN C—short isoform (Δex 16), 5P		9,441		
		24,755	24,755	α _{s1} -CN C—short isoform (Δex 16), 6P	K7DXB9	47,392		
		24,835	24,835	α _{s1} -CN C—short isoform (Δex 16), 7P		7,046		
		24,689	24,689	α _{s1} -CN A—short isoform (Δex 16), 5P		9,748		
		24,768	24,769	α _{s1} -CN A—short isoform (Δex 16), 6P	O97943-2	50,634		
		24,849	24,849	α _{s1} -CN A—short isoform (Δex 16), 7P		6,909		
		VI	28.53	14,430	14,430	α-LAC	P00710	17,797
22,939	n/a			Uncharacterized protein 1 (UP1)	n/a***	2,701		
23,020	n/a			UP1+80Da	n/a	2,489		
23,099	n/a			UP1+160Da	n/a	1,079		
25,646	25,645			α _{s1} -CN C, 6P, splice variant (ΔQ83)		3,501		
25,693	25,693			α _{s1} -CN C, 5P		564		
25,773	25,773			α _{s1} -CN C, 6P		7,880		
25,787	25,787			α _{s1} -CN A, 6P	O97943-1	3,472		
VII	30.05			21,825	21,826	α _{s2} -CN, 7P		552
				21,906	21,906	α _{s2} -CN, 8P	O9794	5,242
				21,984	21,986	α _{s2} -CN, 9P		403
				23,178	n/a	UP1+240Da	n/a	1,256
		22,066	22,066	α _{s2} -CN, 10P	O97944	4,790		
VIII	31.11	21,986	21,986	α _{s2} -CN, 9P		356		
		22,066	22,066	α _{s2} -CN, 10P		4,790		
IX	33.18	22,066	22,066	α _{s2} -CN, 10P		148		
		22,145	22,146	α _{s2} -CN, 11P		1,964		
X	35.05	22,226	22,226	α _{s2} -CN, 12P		894		
		23,046	n/a	Uncharacterized protein 2 (UP2)	n/a	231		
X	37.16	19,143	19,143	PGRP	Q9GK12	7,207		
		23,206	n/a	UP2+160Da	n/a	1,592		
		23,286	n/a	UP2+240Da	n/a	735		
XII	38.09	66,481	66,477	CSA ?	XP_010981066.1	1,096		
			66,491	LPO ?	Q9GJW6			
		66,512	n/a	CSA ? LPO?		2,663		
		67,342	n/a	CSA ? LPO?		1,010		
XIII	40.67	24,746	24,745	β-CN A, 3P, splice variant (ΔQ29)		2,073		
		24,793	24,792	β-CN A, 2P		5,469		
		24,825	24,825	β-CN A, 4P, splice variant (ΔQ29)		9,586		
		24,873	24,872	β-CN A, 3P		10,177		
		24,953	24,953	β-CN A, 4P	A0A077SL35	84,494		
		24,842	24,842	β-CN B, 4P, splice variant (ΔQ29)		10,029		
		24,891	24,890	β-CN B, 3P		10,365		
		24,970	24,971	β-CN B, 4P	Q9TVD0	87,973		
XIV	43.71	23,878	23,878	β-CN A-short isoform (Δ946 Da), 4P, splice variant (ΔQ29)		707		
		23,963	23,958	β-CN A-short isoform (Δ946 Da), 5P, splice variant (ΔQ29)		244		
		23,929	23,926	β-CN A-short isoform (Δ946 Da), 3P		438		
		24,006	24,006	β-CN A-short isoform (Δ946 Da), 4P		9,026		
		23,925	23,925	β-CN A-short isoform (Δ946 Da), 4P, splice variant (ΔQ29)		125		



Mass Spectrometry Analysis
Molecular masses obtained
after deconvolution of
multi-charged ions spectra

αS2-caseins

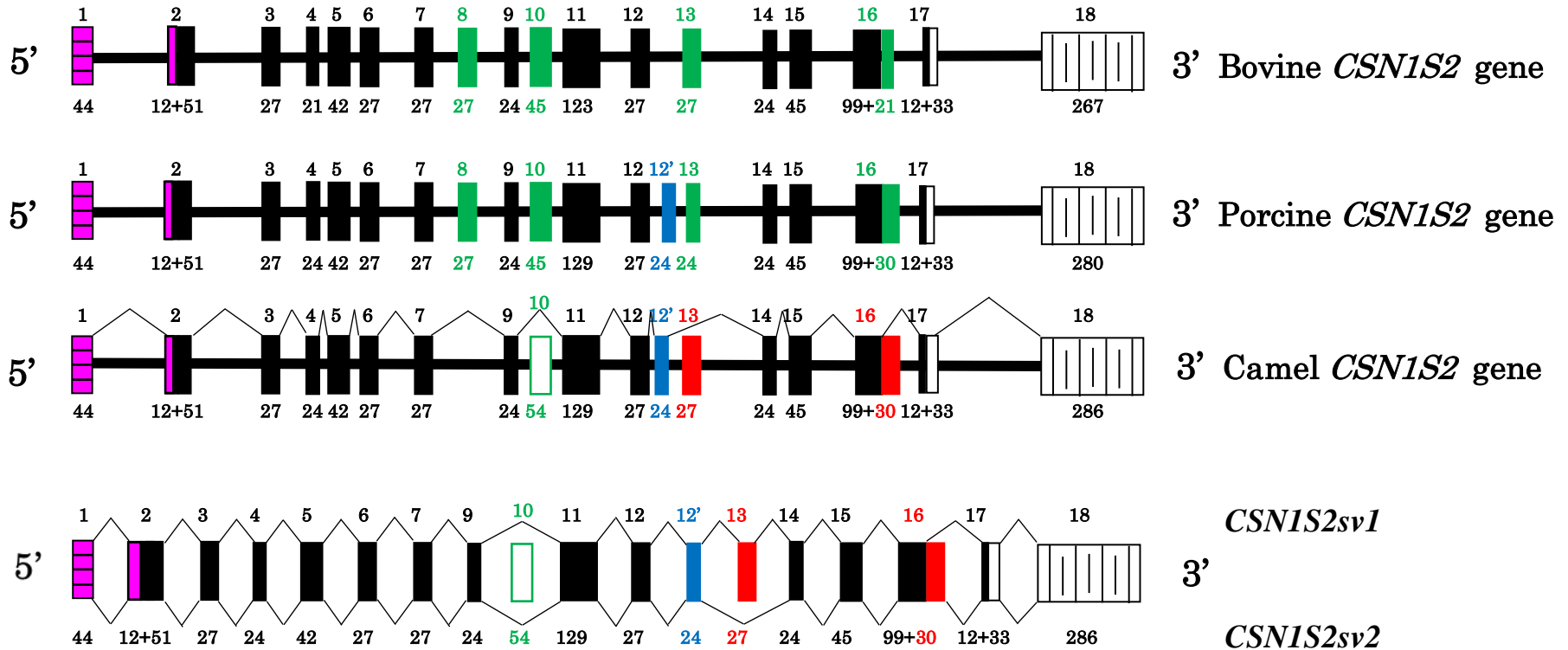


Peak	Ret. Time, min	Observed M_r , Da	Theoretical M_r , Da	Protein description	UniProt/NCBI GenBank Accession number	Intensity
		22,939	n/a	Uncharacterized protein 1 (UPI)	n/a***	2,701
		23,020	n/a	UPI+80Da	n/a	2,489
		23,099	n/a	UPI+160Da	n/a	1,079
		25,646	25,645	α_{s1} -CN C, 6P, splice variant (Δ Q83)		3,501
		25,693	25,693	α_{s1} -CN C, 5P		564
		25,773	25,773	α_{s1} -CN C, 6P		7,880
		25,787	25,787	α_{s1} -CN A, 6P	O97943-1	3,472
VII	30.05	21,825	21,826	α_{s2} -CN, 7P		552
		21,906	21,906	α_{s2} -CN, 8P	O9794	5,242
		21,984	21,986	α_{s2} -CN, 9P		403
		23,178	n/a	UPI+240Da	n/a	1,256
VIII	31.11	21,986	21,986	α_{s2} -CN, 9P	O97944	356
		22,066	22,066	α_{s2} -CN, 10P		4,790
IX	33.18	22,066	22,066	α_{s2} -CN, 10P		148
		22,145	22,146	α_{s2} -CN, 11P		1,964
X	35.05	22,226	22,226	α_{s2} -CN, 12P		894
		23,046	n/a	Uncharacterized protein 2 (UP2)	n/a	231
X	37.16	19,143	19,143	PGRP	Q9GK12	7,207
		23,206	n/a	UP2+160Da	n/a	1,592
		23,286	n/a	UP2+240Da	n/a	735

 α S2-caseins

α S2-caseins

CSN1S2 in camels



Splicing isoforms impacts the C-terminal sequence of camel α_{s2} -casein



	EXON 2	EXON 3	EXON 4	EXON 5	EXON 6	EXON 7	
BOVINE	MKFFIFTCLLAVALA KN	TMEHVSSSE	ESI-ISQE	TYKQEKMAINPSK	ENLCSTFCK	EVVRNANEE	50
LAMA	MKFFIFTCLLAVALA KH	EMDQGSSE	ESINVSQQ	KLK QVKKVAIHPSK	EDICSTFCE	EAVRNIKEV	51
CAMEL	MKFFIFTCLLAVVLA KH	EMDQGSSE	ESINVSQQ	KFKQVKKVAIHPSK	EDICSTFCE	EAVRNIKEV	51
CAMEL SV1	MKFFIFTCLLAVVLA KH	EMDQGSSE	ESINVSQQ	KFKQVKKVAIHPSK	EDICSTFCE	EAVRNIKEV	51
CAMEL SV2	MKFFIFTCLLAVVLA KH	EMDQGSSE	ESINVSQQ	KFKQVKKVAIHPSK	EDICSTFCE	EAVRNIKEV	51
CAMEL SV3	MKFFIFTCLLAVVLA KH	EMDQGSSE	ESINVSQQ	KFKQVKKVAIHPSK	EDICSTFCE	EAVRNIKEV	51
PIG	MKFFIFTCLLAVAF A KH	EMEHVSSSE	ESINISQE	KYKQEKVINHPSK	EDICATSCE	EAVRNIKEV	51

Ryskaliyeva *et al.*, 2019

	EXON 8	EXON 9	EXON 10	EXON 11	
BOVINE	EYSIGSSSE	ESAEVATE	EVKITVDDKHYQKAL	NEINQFYQK--FPQYLQYLYQGPIVLNPWDQVKRNAVPIPTPL	123
LAMA	-----	ESV ^E EVPT E	-----	NKISQFYQKWKFLQYLQALHQGQIVMNPWDQGGK TMVY PFPIPTV	102
CAMEL	-----	ESAEVPT E	-----	NKISQFYQKWKFLQYLQALHQGQIVMNPWDQGGKTRAYPFPIPTV	102
CAMEL SV1	-----	ESAEVPT E	-----	NKISQFYQKWKFLQYLQALHQGQIVMNPWDQGGKTRAYPFPIPTV	102
CAMEL SV2	-----	ESAEVPT E	-----	NKISQFYQKWKFLQYLQALHQGQIVMNPWDQGGKTRAYPFPIPTV	102
CAMEL SV3	-----	ESAEVPT E	-----	NKISQFYQKWKFLQYLQALHQGQIVMNPWDQGGKTRAYPFPIPTV	102
PIG	GYASSSSSE	ESVDIPAE	NVKVTVEDKHYLKQL	EKISQFYQK--FPQYLQALYQAQIVMNPWDQTKTSAYPFPIPTV	124

	EXON 12	EXON 12'	EXON 13	EXON 14	EXON 15	
BOVINE	--NREQLSTSE	-----	ENSKKTVDM	ESTEVFTK	KTKL TEEEKNRNLN FL	164
LAMA	--NTEQLSISE	ESTEVPT E	ENSKKTVDT	ESTEVFTK	KTELTEEEKDHQKFL	151
CAMEL	--NTEQLSISE	ESTEVPT E	-----	ESTEVFTK	KTELTEEEKDHQKFL	146
CAMEL SV1	--NTEQLSISE	ESTEVPT E	ENSKKTVDM	ESTEVFTK	KTELTEEEKDHQKFL	151
CAMEL SV2	--NTEQLSISE	ESTEVPT E	-----	ESTEVFTK	KTELTEEEKDHQKFL	142
CAMEL SV3	--NTEQLSISE	ESTEVPT E	ENSKKTVDM	ESTEVFTK	KTELTEEEKDHQKFL	151
PIG	IQSGEELSTSE	EPVSSSQE	ENT-KTVDM	ESMEEFTK	KTELTEEEKNRKIFL	174

anti-bacterial and antihypertensive activities



	EXON 16	EXON 17	
BOVINE	KKISQRYQKFPALPQYLKTVYQHQA AMKPWIQPKTKV---IPYV	RYL	207
LAMA	NKIYQYYQTFLLWPEYLKTVYQYQKTMTPWNHIK-----	RYF	187
CAMEL	NKIYQYYQTFLLWPEYLKTVYQYQKTMTPWNHIK-----	RYF	178
CAMEL SV1	NKIYQYYQTFLLWPEYLKTVYQYQKTMTPWNHIK-----	RYF	187
CAMEL SV2	NKIYQYYQTFLLWPEYLKTVYQYQKTMTPWNHIK VKAYQIIPNL	RYF	188
CAMEL SV3	NKIYQYYQTFLLWPEYLKTVYQYQKTMTPWNHIK VKAYQIIPNL	RYF	197
PIG	NKIKQYYQKFTWPQYIKTVHQKQKAMKPWNHIK TNSYQIIPNL	RYF	220

Administration of bovine casein-derived peptide prevents cognitive decline in Alzheimer disease model mice

Li-Juan Min¹, Yodai Kobayashi², Masaki Mogi^{1*}, Kana Tsukuda¹, Akio Yamada², Koji Yamauchi², Fumiaki Abe², Jun Iwanami¹, Jin-Zhong Xiao², Masatsugu Horiuchi¹

¹ Department of Molecular Cardiovascular Biology and Pharmacology, Ehime University, Graduate School of Medicine, Shitsukawa, Tohon, Ehime, Japan, ² Morinaga Milk Industry Co., Ltd., Zama, Kanagawa, Japan

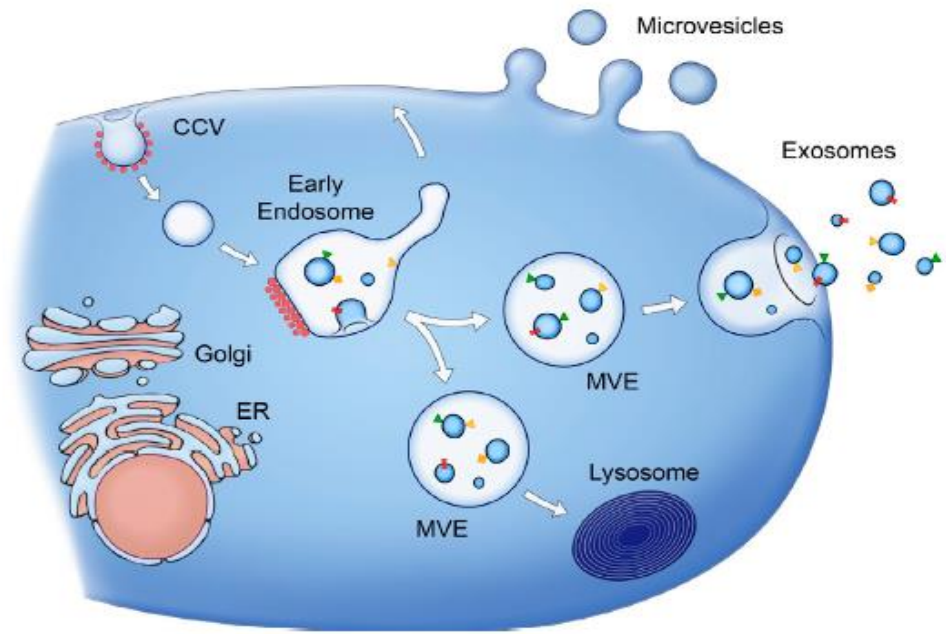
Beside its unique nutritional content breast milk also contains extracellular vesicles and live cells from the mother



Multi Vesicular Bodies released into the extracellular medium by fusion with the cell plasma membrane



Extracellular vesicles
30 and 100 nm



CAMEL MILK-DERIVED EVs: α SOURCE of MILK PROTEINS SO FAR UNEXPLORED

Milk-derived Exosomes and microRNA



Exosomes with Immune Modulatory Features Are Present in Human Breast Milk

Charlotte Admyre, Sara M. Johansson, Khaleda Rahman Qazi, Jan-Jonas Filén, Riitta Lahesmaa, Mikael Norman, Etienne P. A. Neve, Annika Scheynius and Susanne Gabrielsson

J Immunol 2007; 179:1969-1978; ;



This information is current as of September 23, 2019.

Isolation of bovine milk-derived microvesicles carrying mRNAs and microRNAs

Taketoshi Hata^a, Kosuke Murakami^b, Hajime Nakatani^b, Yasunari Yamamoto^c, Tsukasa Matsuda^b, Naohito Aoki^{a,*}

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Biochemical and Biophysical Research Communications 396 (2010) 528–533



MicroRNAs: Milk's epigenetic regulators

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Best Practice & Research Clinical Endocrinology & Metabolism 31 (2017) 427–442



Human Milk MicroRNAs/Exosomes: Composition and Biological Effects

Bo Lönnerdal

Donovan SM, German JB, Lönnerdal B, Lucas A (eds): Human Milk: Composition, Clinical Benefits and Future Opportunities. Nestlé Nutr Inst Workshop Ser, vol 90, pp 83–92, (DOI: 10.1159/000490297)
Nestlé Nutrition Institute, Switzerland/S. Karger AG., Basel, © 2019

Extracellular vesicles in camel milk

Comprehensive proteomic analysis of camel milk-derived extracellular vesicles

¹*A. Ryskaliyeva, ²Z. Krupova, ³C. Henry, ⁴B. Faye, ⁵G. Konuspayeva, ¹P. Martin

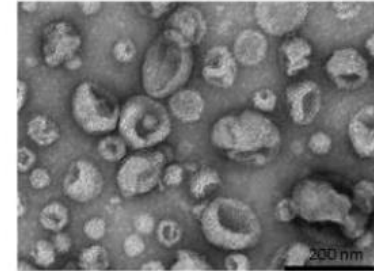
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Paris Sud-Ouest, Université Paris-Saclay, France

⁴CIRAD, UMR SELMET, France

⁵RPE Co Antigen Ltd., Kazakhstan



International Journal of Biology and Chemistry 12, № 2, 93 (2019)



Therapeutic potential of camel milk exosomes against HepaRG cells with potent apoptotic, anti-inflammatory, and anti-angiogenesis effects for colostrum exosomes

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Yousef M. Hawsawi^{d,e}, Othman R. Alzahrani^{f,g}, Abdulrahman Algarni^h, Khaled A. Kahilo^a,
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Therapeutic Effect of Camel Milk and Its Exosomes on MCF7 Cells In Vitro and In Vivo

Integrative Cancer Therapies

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and Sana A. AISadrah, PhD⁴

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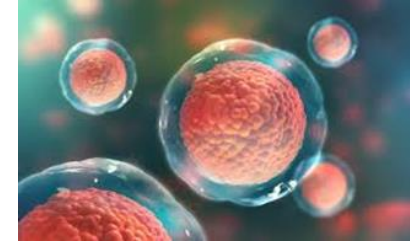
Breast milk stem cells transfer to offspring

SCIENTIFIC REPORTS

SCIENTIFIC REPORTS | (2018) 8:14289 | DOI:10.1038/s41598-018-32715-5

Transfer and Integration of Breast Milk Stem Cells to the Brain of Suckling Pups

Mehmet Şerif Aydın¹, Esra Nur Yiğit¹, Emre Vatandaşlar¹, Ender Erdoğan² & Gürkan Öztürk^{1,3}



Milk SC reach the brain where they turn into functioning cells

Accepted: 28 November 2017

DOI: 10.1111/pai.12841




REVIEW ARTICLE

WILEY

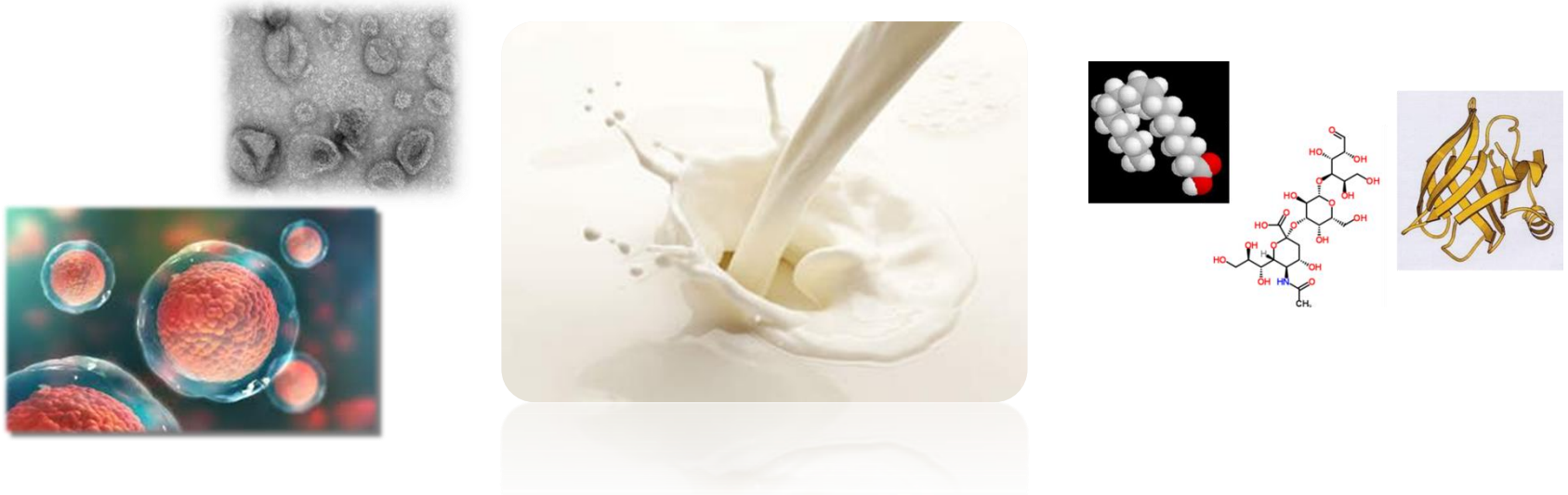


Breastmilk cell trafficking induces microchimerism-mediated immune system maturation in the infant

Jean-Pierre Molès¹ | Edouard Tuaillon^{1,2} | Chipepo Kankasa³ | Anne-Sophie Bedin¹ | Nicolas Nagot^{1,2} | Arnaud Marchant⁴ | Joann M. McDermid⁵ | Philippe Van de Perre^{1,2} 

Breast-milk: the postnatal maternal blood through which a multitude of active soluble and cellular factors are delivered to the offspring

Conclusions



Milk has to be considered as a sophisticated nutrient but also as a **communication system delivering** a multitude of active soluble and cellular factors orchestrating early development of the offspring

Differential splicing is an effective way to **increase** molecular diversity and possibly **the repertoire of bioactive peptides** encrypted in milk proteins

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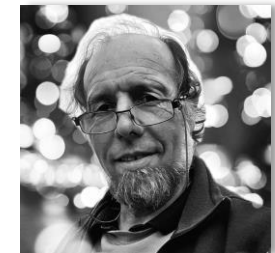


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